



Euskal Herriko Unibertsitatea
Universidad del País Vasco

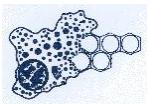


ZTF-FCT
Zientzia eta Teknologia Fakultatea
Facultad de Ciencia y Tecnología

Universidad
del País Vasco
Euskal Herriko
Unibertsitatea



Cell Biology in Environmental Toxicology (CBET) consolidated research group: Nanotoxicology research line



Contact information:

Prof. Dr. Miren P. Cajaraville

Director Consolidated Research Group "Cell Biology in Environmental Toxicology CBET"
Department of Zoology and Animal Cell Biology
Faculty of Science and Technology
University of the Basque Country UPV/EHU
Sarriena Auzoa z/g, E-48940 Leioa, Basque Country (Spain)

Research Centre for Experimental Marine Biology and Biotechnology
Plentzia Marine Station PiE-UPV/EHU,
Areatza z/g, E-48620 Plentzia, Basque Country (Spain)

e-mail: miren.p.cajaraville@ehu.eus
Tel: + 34 94 6012697 (office) + 34 94 6015416 (lab)
Fax: + 34 94 6013500
Research Group CBET:
www.ehu.eus/web/cellbiologyinenvironmentaltoxicology
Plentzia Marine Station PiE-UPV/EHU: www.ehu.es/PIE

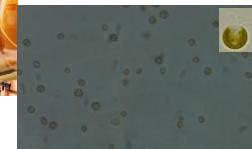
Presentation of the CBET group

Since 1985 the group of "Cell Biology in Environmental Toxicology" of the University of the Basque Country develops research and teaching activities within the Environmental Toxicology field (both aquatic and terrestrial ecosystems). The group, recognized since 2001 as consolidated research group, is composed of 10 university lecturers and researchers, 4 postdoc researchers and 12 PhD students. The group's lab contains facilities for histology, light, fluorescence and electron microscopy, image analysis, cell fractionation and biochemistry, cell culture and molecular biology. The group has aquaria facilities for development of large experiments and mesocosm experiments and a zebrafish facility at the Faculty of Science and Technology and at the Research Centre for Experimental Marine Biology and Biotechnology, Plentzia Marine Station PiE-UPV/EHU. The group coordinates three Masters (two recognized with the Erasmus Mundus label) and two Doctoral Programmes (Environmental Contamination and Toxicology, Marine Environment and Resources) and offers several international postgraduate courses.

The main field of expertise of the group is in development of early warning cell and molecular biomarkers of pollution exposure and effects, and in toxicity testing of environmental pollutants using standard and novel toxicity tests in cells *in vitro*, algae, invertebrates and vertebrates.

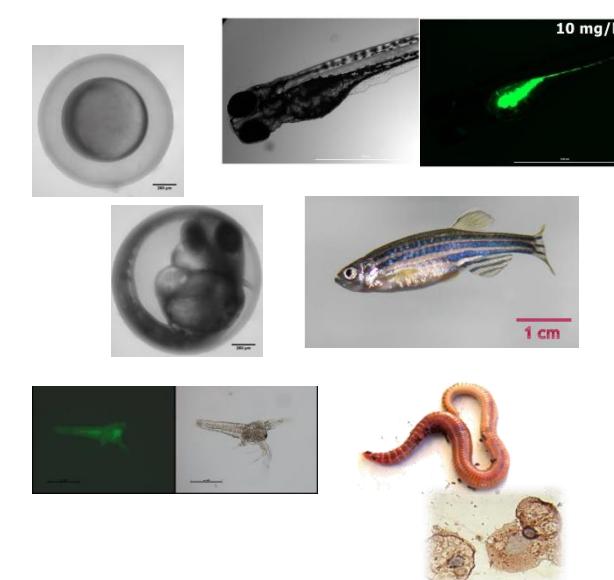
Main research lines of the group are:

- Ecosystem health assessment
- Effects of petroleum hydrocarbons, oil spills and produced water
- Endocrine disruption
- Toxicogenomics
- Chemical carcinogenesis
- Nanotoxicology



R+D+I services offered

- Scientific and technical advising. Areas: pollution, accidental spills, environmental toxicology, use of biomarkers, cell culture and *in vitro* systems, microscopy, molecular, cell and tissue organization of animals.
- Scientific education: Masters, postgraduate courses, specialization courses, PhD programmes, laboratory training and internships.
- Assessment of environmental quality (marine, estuarine, and terrestrial environments).
- Monitoring programmes of environmental pollution.
- Toxicity testing (REACH legislation).
- Determination of biological activity of chemicals.
- Histopathological diagnosis of fish and shellfish diseases.
- Processing of samples for histology, light and electron microscopy, cryotechniques.
- Quantification of levels of specific proteins, enzyme activities, gene expression, immunolabelling, cell structure.
- Establishment and maintenance of experimental animals. Marine aquaria. Zebrafish facility. Terrarium. Facilities to run chemical exposure experiments.



Nanotoxicology research line

In the Nanotoxicology field, we have been involved in several projects including NanoReTox (EU 7th FP, 2008-2012), NanoCancer (Spanish MEC, 2010-2012), NanoSilverOmics (Spanish MINECO, 2013-2015), Enter (EU Cost Action, 2013-2017), NanoToxT (Basque Gov Saitotek, 2013-2014) and NanoGune (Basque Gov Etortek, 2014-2015). Currently we are carrying out the project NanoCarrierERA-NACE (Spanish MINECO, 2016-2020) and we are part of the National Network of Excellence in Nanotechnology and Food (Spanish MINECO, 2017-2019).

The **main goals** of the Nanotoxicology research line are:

- Understanding the behaviour of nanomaterials in different exposure media
- Determination of uptake, bioaccumulation, bioavailability, fate, and cellular and tissue distribution of nanomaterials in organisms.
- Identification of mechanisms of action and adverse effects of nanomaterials in organisms
- Toxicity profiling of nanomaterials
- Risk assessment of nanomaterials in aquatic and terrestrial ecosystems

Selected publications

DUROUDIER, N; KATSUMITI, A; MIKOLACZYK, M; SCHÄFER, J; BILBAO, E; CAJARAVILLE, MP. 2019. Dietary exposure of mussels to PVP/PEI coated Ag nanoparticles causes Ag accumulation in adults and abnormal embryo development in their offspring. *Science of the Total Environment*, 655: 48-60.

DUROUDIER, N; CARDOSO, C; MEHENNAOUI, K; MIKOLACZYK, M; SCHÄFER, J; GUTLEB, AC; GIAMBERINI, L; BEBIANNO, MJ, BILBAO, E; CAJARAVILLE, MP. 2019. Changes in protein expression in mussels *Mytilus galloprovincialis* dietarily exposed to PVP/PEI coated silver nanoparticles at different seasons. *Aquatic Toxicology*, 210: 56-68.

DUROUDIER, N; MARKAIDE, P; CAJARAVILLE, MP; BILBAO, E. 2019. Season influences the transcriptomic effects of dietary exposure to PVP/PEI coated silver nanoparticles on mussels *Mytilus galloprovincialis*. *Comparative Biochemistry and Physiology, Ser C*, 222: 19-30.

GARCIA-VELASCO, N; GANDARIASBEITIA, M; IRIZAR, A; SOTO, M. 2016. Uptake route and resulting toxicity of silver nanoparticles in *Eisenia fetida* earthworm exposed through Standard OECD Tests. *Ecotoxicology*, 25: 1543-1555.

GARCIA-VELASCO, N; PEÑA-CEARRA, A; BILBAO, E; ZALDIBAR, B; SOTO, M. 2017. Integrative assessment of the effects produced by Ag nanoparticles at different levels of biological complexity in *Eisenia fetida* maintained in two standard soils (OECD and LUFA 2.3). *Chemosphere*, 181: 747-758.

GONZÁLEZ-SOTO, N; HATFIELD, J; KATSUMITI, A; DUROUDIER, N; LACAVE, JM; BILBAO, E; ORBEA, A; NAVARRO, E; CAJARAVILLE, MP. 2019. Impacts of dietary exposure to different sized polystyrene microplastics alone and with sorbed benzo[a]pyrene on biomarkers and whole organism responses in mussels *Mytilus galloprovincialis*. *Science of the Total Environment*, doi: 10.1016/j.scitotenv.2019.05.161.

JIMENO-ROMERO, A; BILBAO, E; IZAGIRRE, U; CAJARAVILLE, MP; MARIGÓMEZ, I; SOTO, M. 2017. Digestive cell lysosomes as main targets for Ag accumulation and toxicity in marine mussels, *Mytilus galloprovincialis*, exposed to maltose-stabilised Ag nanoparticles of different sizes. *Nanotoxicology*, 11: 168-183.

JIMENO-ROMERO, A; IZAGIRRE, U; GILLILAND, D; WARLEY, A; CAJARAVILLE, MP; MARIGOMEZ, I; SOTO, M. 2017. Lysosomal responses to different gold forms (nanoparticles, aqueous, bulk) in mussel digestive cells: a trade-off between the toxicity of the capping agent and form, size and exposure concentration. *Nanotoxicology*, 11: 1-32.

JIMENO-ROMERO, A; BILBAO, E; VALSAMÍ-JONES, E.; CAJARAVILLE, MP; SOTO, M; MARIGÓMEZ, I. 2019. Bioaccumulation, tissue and cell distribution, biomarkers and toxicopathic effects of CdS quantum dots in mussels, *Mytilus galloprovincialis*. *Ecotoxicology and Environmental Safety*, 167: 288-300.

JIMENO-ROMERO, A; ORON, M; CAJARAVILLE, MP; MARIGÓMEZ, I; SOTO, M. 2016. Nanoparticle size and combined toxicity of TiO₂ and DSLS (surfactant) contribute to lysosomal responses in digestive cells of mussels exposed to TiO₂ nanoparticles. *Nanotoxicology*, 10: 1168-76.

KATSUMITI, A; AROSTEGUI, I; ORON, M; GILLILAND, D; VALSAMÍ-JONES, E; CAJARAVILLE, MP. 2016. Cytotoxicity of Au, ZnO and SiO₂ nanoparticles using in vitro assays with mussel hemocytes and gill cells: relevance of size, shape and additives. *Nanotoxicology*, 10: 185-193.

KATSUMITI, A; BERHANU, D; HOWARD, KT; AROSTEGUI, I; ORON, M; REIP, P; VALSAMÍ-JONES, E; CAJARAVILLE, MP. 2015. Cytotoxicity of TiO₂ nanoparticles to mussel hemocytes and gill cells in vitro: influence of synthesis method, crystalline structure, size and additive. *Nanotoxicology*, 9: 543-553.

KATSUMITI, A; CAJARAVILLE, MP. 2019. Chapter 9: In vitro testing. In: *Ecotoxicology of nanoparticles in aquatic systems*. J BLASCO, I CORSI (eds), Science Publishers (CRC Press/ Taylor & Francis Group), *in press*.

KATSUMITI, A; THORLEY, A; AROSTEGUI, I; REIP, P; VALSAMÍ-JONES, E; TETLEY, TD; CAJARAVILLE, MP. 2018. Cytotoxicity and cellular mechanisms of toxicity of CuO NPs in mussel cells in vitro and comparative sensitivity with human cells. *Toxicology in Vitro*, 48: 146-158.

KATSUMITI, A; GILLILAND, D; AROSTEGUI, I; CAJARAVILLE, MP. 2014. Cytotoxicity and cellular mechanisms involved in the toxicity of CdS quantum dots in hemocytes and gill cells of the mussel *Mytilus galloprovincialis*. *Aquatic Toxicology*, 153: 39-52.

KATSUMITI, A; GILLILAND, D; AROSTEGUI, I; CAJARAVILLE, MP. 2015. Mechanisms of toxicity of Ag nanoparticles in comparison to bulk and ionic Ag on mussel hemocytes and gill cells. *PLOS One*, 10(6): e0129039. doi: 10.1371/journal.pone.0129039, 1-30.

KATSUMITI, A; TOMOVSKA, R; CAJARAVILLE, MP. 2017. Intracellular localization and toxicity of graphene oxide and reduced graphene oxide nanoplatelets to mussel hemocytes in vitro. *Aquatic Toxicology*, 188: 138-147.

LACAVE, JM; RETUERTO, A; VICARIO-PARÉS, U; GILLILAND, D; ORON, M; CAJARAVILLE, MP; ORBEA, A. 2016. Effects of metal bearing nanoparticles (Ag, Au, CdS, ZnO, SiO₂) on developing zebrafish embryos. *Nanotechnology*, 27: 325102-325116.

LACAVE, JM; FANJUL, A; BILBAO, E; GUTIERREZ, N; BARRIO, I; AROSTEGUI, I; CAJARAVILLE, MP; ORBEA, A. 2017. Acute toxicity, bioaccumulation and effects of dietary transfer of silver from brine shrimps exposed to PVP/PEI-coated silver nanoparticles to zebrafish. *Comparative Biochemistry and Physiology - Part C: Toxicology & Pharmacology*, 199: 69-80.

LACAVE, JM; VICARIO-PARÉS, U; BILBAO, E; GILLILAND, D; MURA, F; DINI, L; CAJARAVILLE, MP; ORBEA, A. 2018. Waterborne exposure of adult zebrafish to silver nanoparticles and to ionic silver results in differential silver accumulation and effects at cellular and molecular levels. *Science of the Total Environment*, 642: 1209-1220.

ORBEA, A; VICARIO-PARÉS, U; LACAVE, JM; CAJARAVILLE, MP. 2015. El pez cebra como modelo experimental en Nanotoxicología. *Revista de la Sociedad Española para las Ciencias del Animal de Laboratorio SECAL*, 64: 41-44.

ORBEA, A; GONZÁLEZ-SOTO, N; LACAVE, JM; BARRIO, I; CAJARAVILLE, MP. 2017. Developmental and reproductive toxicity of PVP/PEI-coated silver nanoparticles to zebrafish. *Comparative Biochemistry and Physiology, Ser C*, 199: 59-68.

RUIZ, P; KATSUMITI, A; NIETO, JA; BORI, J; REIP, P; ORBEA, A; CAJARAVILLE, MP. 2015. Short-term effects on antioxidant enzymes and long-term genotoxic and carcinogenic potential of CuO nanoparticles in mussels *Mytilus galloprovincialis*. *Marine Environmental Research*, 111: 107-120.

VICARIO-PARÉS, U; CASTAÑAGA, L; LACAVE, JM; ORON, M; REIP, P; BERHANU, D; VALSAMÍ-JONES, E; CAJARAVILLE, MP; ORBEA, A. 2014. Comparative toxicity of metal oxide nanoparticles (CuO, ZnO, TiO₂) to developing zebrafish embryos. *J Nanoparticle Research*, 16: 2550-2566.

VICARIO-PARÉS, U; LACAVE, JM; REIP, P; CAJARAVILLE, M.P.; ORBEA A. 2017. Cellular and molecular responses of adult zebrafish after exposure to CuO nanoparticles or ionic copper. *Ecotoxicology*, 27: 89-101.